axol

Human Mammary Epithelial Cells

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Catalog. No.	Product Name	Format	Short-term Storage	Long-term Storage	Thawing Instructions
ax3512	Mammary Epithelial Cells (Male)	500,000 cells/vial	Liquid Nitrogen	Liquid Nitrogen	See below
ax3513	Mammary Epithelial Cells (Female)	500,000 cells/vial	Liquid Nitrogen	Liquid Nitrogen	See below
ax3537	Mammary Epithelial Cell Culture Medium	500 mL	Store at 4°C for up to 1 month	Aliquot and store at -20°C for up to 6 months	Thaw at 4°C or at room temperature

Lot-specific information such as donor details and passage number are stated in the Certificate of Analysis for each product.

Recommendations

Important!

Always count the number of viable cells after thawing.

- Recommended culture vessel coating: Not required
- Recommended cell culture medium: **Mammary Epithelial Cell Culture Medium**
- Recommended seeding density: **5,000 viable cells/cm²**
- Recommended centrifugation speed: **200 x *g*** for **5 minutes**

Thawing and Plating

- Transfer the vial of cells from liquid nitrogen storage with the vial buried in dry ice. Remove the vial from dry ice and transfer it immediately to a **37°C** water bath.
- Thaw the cells quickly in a **37°C** water bath until just prior to complete thawing.
- Wipe the outside of the vial with 70% ethanol.
- Gently resuspend the cells and take an aliquot to perform a cell count.
- **Immediately after thawing**, slowly dilute the cells into the required volume of pre-warmed **Mammary Epithelial Cell Culture Medium** (must be at least **10 mL** so that the concentration of DMSO is less than 1%).
- Rinse the cryovial with **1 mL** of **Mammary Epithelial Cell Culture Medium** to ensure all of the cells are transferred.
- Seed cells into the culture vessel at the recommended seeding density of **5,000 viable cells/cm²**.
- Incubate the cells at **37°C**, **5% CO_2** in a humidified incubator.
- Once the cells have attached (after 6-24 hours), replace the culture medium with fresh, pre-warmed **Mammary Epithelial Cell Culture Medium**.
- Frequency of media changes: **Every 2 days**

Passaging

- Passage when the culture reaches: **95-100% confluent**
- Recommended passaging reagent: **Trypsin-EDTA**
- After adding passaging reagent, incubate the cells for **5 minutes** at **37°C**. Observe the cells at regular intervals for detachment from the culture vessel.
- Once the cells have detached, dilute out the trypsin with pre-warmed **Mammary Epithelial Cell Culture Medium**.
- Centrifuge the cells at **200 x *g*** for **5 minutes**.
- Remove the supernatant and resuspend the cell pellet in **1-2 mL** of pre-warmed **Mammary Epithelial Cell Culture Medium**.
- Perform a cell count to determine the number of viable cells.
- Dilute the cells into the required volume of pre-warmed **Mammary Epithelial Cell Culture Medium**.
- Seed cells into the culture vessel at the recommended seeding density of **5,000 viable cells/cm^2**.
- Incubate the cells at **37°C**, **5% CO_2** in a humidified incubator.

Important!

It is recommended that low passage **Mammary Epithelial Cells** are used for endpoint assays since the proportion of luminal cells may decrease upon passaging and the proportion of basal cells may increase upon passaging.

Usage Statement

Our products are intended for research use only and are not to be used for any other purpose, which includes but is not limited to, unauthorized commercial uses, *in vitro* diagnostic uses, *ex vivo* or *in vivo* therapeutic uses or any type of consumption or application to humans.

Got any questions? Need help with the protocol?
Contact Axol Technical Support at **support@axolbio.com**
International phone **+44-1223-751-051**
US phone **+1-800-678-AXOL (2965)**

Notes

Notes

Address

Axol Bioscience Limited | Suite 3 | The Science Village |
Chesterford Research Park | Little Chesterford | Cambridgeshire | CB10 1XL

International phone

+44-1223-751-051

US phone

+1-800-678-AXOL (2965)

Email

support@axolbio.com

Web

www.axolbio.com