axol

Human Fibroblasts

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Catalog. No.	Product Name	Format	Short-term Storage	Long-term Storage	Thawing Instructions
ax3010 ax3011 ax3012 ax3013 ax3014 ax3015 ax3016 ax3017 ax3018 ax3019 ax3020 ax3021 ax3022 ax3023 ax3024 ax3025 ax3027 ax3030 ax3031 ax3032 ax3033 ax3034 ax3035 ax3036 ax3037 ax3038 ax3039 ax3040	Human Fibroblasts	500,000 cells/vial	Liquid Nitrogen	Liquid Nitrogen	See below
ax3045	Fibroblast Plating & Growth Medium	500 mL	4°C for 1 month	-20°C for 6 months	Thaw at 4°C or RT

Lot-specific information such as donor details and passage number are stated in the Certificate of Analysis for each product.

Recommendations

Important!

Always count the number of viable cells after thawing.

- Recommended culture vessel coating: Not required
- Recommended cell culture medium: **Fibroblast Plating & Growth Medium**
- Recommended seeding density: **4,000 viable cells/cm²**
- Recommended centrifugation speed: **200 x *g*** for **5 minutes**

Thawing and Plating

- Transfer the vial of cells from liquid nitrogen storage with the vial buried in dry ice. Remove the vial from dry ice and transfer it immediately to a **37°C** water bath.
- Thaw the cells quickly in a **37°C** water bath until just prior to complete thawing.
- Wipe the outside of the vial with 70% ethanol.

- Gently resuspend the cells and take an aliquot to perform a cell count.
- **Immediately after thawing**, slowly dilute the cells into the required volume of pre-warmed **Fibroblast Plating & Growth Medium** (must be at least **10 mL** so that the concentration of DMSO is less than 1%).
- Rinse the cryovial with **1 mL** of **Fibroblast Plating & Growth Medium** to ensure all of the cells are transferred.
- Seed cells into the culture vessel at the recommended seeding density of **4,000 viable cells/cm²**.
- Incubate the cells at **37°C**, **5% CO_2** in a humidified incubator.
- Once the cells have attached (after 6-24 hours), replace the culture medium with fresh pre-warmed **Fibroblast Plating & Growth Medium**.
- Frequency of media changes: **Every 3 days**

Passaging

- Passage when the culture reaches: **80-90% confluent**
- Recommended passaging reagent: **Trypsin-EDTA**
- After adding passaging reagent, incubate the cells for **5 minutes** at **37°C**. Observe the cells at regular intervals for detachment from the culture vessel.
- Once the cells have detached, dilute out the trypsin with **Fibroblast Plating & Growth Medium** and centrifuge the cells at **200 x *g*** for **5 minutes**.
- Remove the supernatant and resuspend in **1-2 mL** of pre-warmed **Fibroblast Plating & Growth Medium**.
- Perform a cell count to determine the number of viable cells.
- Dilute the cells into the required volume of pre-warmed **Fibroblast Plating & Growth Medium**.
- Seed cells into the culture vessel at the recommended seeding density of **4,000 viable cells/cm²**.
- Incubate the cells at **37°C**, **5% CO_2** in a humidified incubator.

Usage Statement

Our products are intended for research use only and are not to be used for any other purpose, which includes but is not limited to, unauthorized commercial uses, *in vitro* diagnostic uses, *ex vivo* or *in vivo* therapeutic uses or any type of consumption or application to humans.

Got any questions? Need help with the protocol?
Contact Axol Technical Support at **support@axolbio.com**
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Notes

Notes

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